

**Mixed Phenols and  
Phenates by LC- UV  
AOAC International  
Collaborative Study**

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# US EPA Pesticide Registration Guidelines:

- The EPA requires a suitable analytical method for all registered pesticide active ingredients in technical materials and if requested, end use products
- Enforcement Analytical methods are not required to have a Single Laboratory Validation (SLV) for EPA registration.

# US EPA Enforcement Concerns:

- Disagreements over method used have occurred between registrant and FIFRA (Federal Insecticide, Fungicide, and Rodenticide Act) laboratory
- Government laboratories need accurate, reliable and rugged methods for their enforcement work.
- The chlorinated phenols, used in many hospital disinfectants had not been collaboratively studied.

# Preliminary work:

- We considered two methods for potential collaborative study, a GC method and an LC method
- A pilot study was conducted for both methods using the same 8 commercial disinfectant samples analyzed by 6 laboratories.
- All laboratories returned data using the LC method, but only 2 laboratories returned data on the GC method.

- From the results and comments from the pilot study, a single laboratory validation for the LC method was conducted by Tom Phillips, Maryland Department of Agriculture to determine the accuracy and repeatability (within lab).
- The SLV was published in the Journal of the AOAC International (T.Phillips, A. Burns, 2010)

# Study Design:

- We wanted a minimum of ten laboratories from government, industry and academia to collaboratively study the proposed method.
- We wanted a minimum of five materials encompassing range of % actives and variety of inert ingredients.

# Mixed Phenols/Phenates Collaborative Study Design

- **Originally had 19 laboratories sign up for the study**
- **1 test sample to ensure analyst familiarity with the method and check out the LC system**
- **7 Samples selected reflect as much variability as possible:**
  - **Low levels**
  - **High levels**
  - **Salts (phenates)**
  - **Other active ingredients**

# Phenols/Phenates Examined:

- **Opp:** Ortho phenyl phenol / phenate
- **Obpcp:** ortho benzyl parachloro-phenol/phenate
- **Ptap:** para tertiary amyl phenol / phenate
- These are the top three registered phenols and present as single or multi-active ingredient in 89 different currently registered products.



# Participating Laboratories:

- Iowa Department of Agriculture and Land Stewardship
- Robert Wesleyan College
- North Carolina Department of Agriculture and Consumer Services
- Georgia Department of Agriculture
- Steris Corporation
- Lonza Corporation

- Clorox Corporation
- Florida Department of Agriculture/  
Environmental Services
- US Environmental Protection Agency
- Dow AgroSciences
- Silliker Laboratories
- Kansas Department of Agriculture
- Maryland Department of Agriculture

# Sample Table

	Content expressed as per cent phenol			
Sample no.	Opp	Ptap	Obpcp	Notes:
1	2.50	2.20	4.00	K salt
2	5.13	<b>3.47</b>	6.84	Na salt
3	<b>0.04</b>	<b>0.07</b>	<b>0.08</b>	
4	4.90	2.50	<b>10.10</b>	
5	4.02	1.20	4.90	
6	2.35	3.47	3.80	
7	<b>10.00</b>	2.00	8.50	

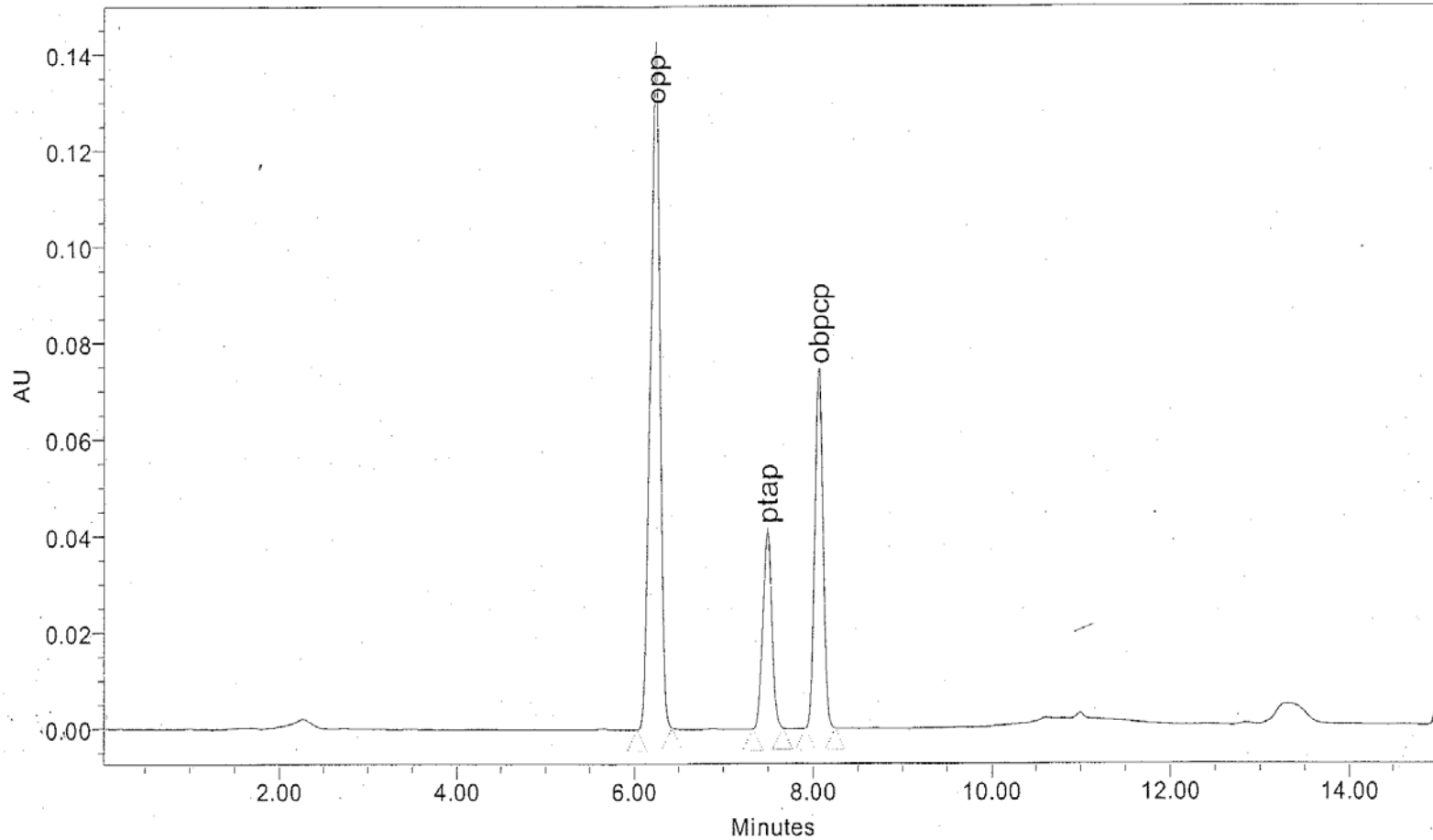
# LC Columns Used:

- Phenomenex Luna C18(2)
- Waters  $\mu$ Bondapak ODS (300x3.9 mm)
- Waters Novapak ODS (200x 4.0 mm)
- Column Engineering Inertsil
- Keystone Scientific Betasil ODS
- Agilent Zorbax Eclipse XDB-ODS
- Phenomenex Kinetix ODS (100x 4.6 mm)
- Whatman Partisphere ODS (250x 4.6 mm)

# Method Summary:

- Samples are extracted with acidified methanol
- LC is run with a gradient mobile phase of acidified water/ acetonitrile
- Detector is UV at 285 nm

# Typical chromatogram



# **Samples were tested as blind duplicates**

- Data was presented by analyte:**
- % concentration found**
- Stats for outliers- Cochran and Single Grubbs run**
- Predicted relative standard deviation**
- Relative standard deviation found**
- HorRat was determined for each analyte of interest**

Analyte	No. of Labs (outliers)	Mean % <sup>a</sup>	$s_r$ <sup>b</sup>	$s_R$ <sup>c</sup>	$RSD_r$ <sup>d</sup>	$RSD_R$ <sup>e</sup>	$r$ <sup>f</sup>	$R$ <sup>g</sup>	HorRat	Outlier Labs <sup>h</sup>
<b>OPP</b>	<b>13(1)</b>	<b>4.840</b>	<b>0.0719</b>	<b>0.2791</b>	<b>1.49</b>	<b>5.77</b>	<b>0.201</b>	<b>0.781</b>	<b>1.83</b>	<b>2-C</b>
	<b>13(1)</b>	<b>0.047</b>	<b>0.0005</b>	<b>0.0025</b>	<b>0.98</b>	<b>5.43</b>	<b>0.001</b>	<b>0.007</b>	<b>0.86</b>	<b>2-C</b>
	<b>14(0)</b>	<b>2.463</b>	<b>0.0751</b>	<b>0.1551</b>	<b>3.05</b>	<b>6.30</b>	<b>0.210</b>	<b>0.434</b>	<b>1.80</b>	
	<b>13(1)</b>	<b>3.649</b>	<b>0.0698</b>	<b>0.2196</b>	<b>1.91</b>	<b>6.02</b>	<b>0.195</b>	<b>0.615</b>	<b>1.83</b>	<b>16-C</b>
	<b>14(0)</b>	<b>9.953</b>	<b>0.1166</b>	<b>0.6123</b>	<b>1.17</b>	<b>6.15</b>	<b>0.326</b>	<b>1.714</b>	<b>2.17</b>	

<sup>a</sup> Weight percent average of the blind duplicate pair

<sup>b</sup>  $s_r$  = Standard deviation for repeatability (within laboratory).

<sup>c</sup>  $s_R$  = Standard deviation for reproducibility (among laboratories).

<sup>d</sup>  $RSD_r$  = Relative standard deviation for repeatability.

<sup>e</sup>  $RSD_R$  = Relative standard deviation for reproducibility.

<sup>f</sup>  $r = 2.8 * s_r$ .

<sup>g</sup>  $R = 2.8 * s_R$ .

<sup>h</sup> C= Cochran outlier; SG = single Grubbs outlier.



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<b>PTAP</b>	13(1)	2.551	0.0397	0.1730	1.56	6.78	0.111	0.484	1.95	2-C
	12(2)	0.054	0.0009	0.0050	1.12	5.95	0.002	0.014	1.03	2,4-C
	14(0)	3.959	0.0870	0.2103	2.20	5.31	0.244	0.589	1.63	
	14(0)	1.143	0.0389	0.0891	3.40	7.80	0.109	0.249	1.99	18-SG
	13(1)	1.961	0.0467	0.1074	2.38	5.48	0.131	0.301	1.52	18-SG

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OBPCP	14(0)	10.150	0.3267	0.5469	3.22	5.39	0.915	1.53	1.91	
	13(1)	0.095	0.0014	0.0053	1.49	5.57	0.004	0.015	0.98	2-C
	13(1)	4.276	0.0598	0.2356	1.40	5.51	0.167	0.660	1.71	19-C
	13(1)	4.855	0.0920	0.2709	1.90	5.58	0.258	0.758	1.77	16-C
	14(0)	8.815	0.1142	0.4845	1.30	5.50	0.320	1.357	1.91	

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<b>Results for the salt forms of OPP, PTAP, and OBPCP</b>										
OPP	14(0)	2.614	0.0520	0.1495	1.99	5.72	0.146	0.419	1.65	
	14(0)	6.123	0.0859	0.3835	1.40	6.26	0.241	1.074	2.06	
PTAP	13(1)	2.150	0.0540	0.1182	2.51	5.50	0.151	0.331	1.54	8-SG
	14(0)	3.933	0.0734	0.2508	1.87	6.38	0.206	0.702	1.96	
OBPCP	14(0)	4.471	0.1107	0.3877	2.47	8.67	0.310	1.086	2.72	
	13(1)	8.252	0.1037	0.4942	1.26	5.99	0.290	1.384	2.06	12-C

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# Results

- Results were acceptable. There were 26 outliers in 588 data points generated (4.42%)
- Performance of the method compared favorably to the SLV as well.
- AOAC International granted First Action in 2011 (AOAC 2011.26)

# Acknowledgements:

- **Tom Phillips for conducting the Single Laboratory Validation and being a co-study director**
- **US EPA for funding this collaborative study**
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# Questions?

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Thank You!